

with  $\text{NH}_3$ , at  $R_f$  values of 4–20 ( $\text{H}_2\text{O}$ ) and 16–50 ( $\text{HOAc}$ ). The species with C-glycosylxanthones were subjected to 2D-PC in TBA and 15%  $\text{HOAc}$  [9]. The compounds were eluted in 80%  $\text{MeOH}$  for  $R_f$  comparison in TBA, BAW, 15%  $\text{HOAc}$  and  $\text{H}_2\text{O}$  with authentic mangiferin and isomangiferin from *Asplenium montanum* [10].

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## FLAVONE C-GLYCOSIDES OF *PHORADENDRON TOMENTOSUM* FROM DIFFERENT HOST TREES

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**Key Word Index**—*Phoradendron tomentosum*, mistletoes, Loranaceae, flavones, apigenin mono- and di-C-glycosides

**Abstract**—Apigenin, three known apigenin C-glycosides, vitexin, schaftoside and isoschaftoside together with apigenin 4'-O-glucoside have been identified in leaves of *Phoradendron tomentosum* growing on different hosts

Mistletoes are semi-parasitic plants which in modern botanical taxonomy are classified into two families, the Loranaceae and Viscaceae [1]. In an earlier paper Becker and Exner [2] reported the isolation of eight flavonoids from *Viscum album* all of which were methylated quercetin derivatives. The present paper describes the characterization of apigenin C-glycosides from plants of *P. tomentosum* (DC.) Gray, growing on three different host trees: *Ulmus crassifolia* Nutt., *Prosopis glandulosa* Torr. and *Celtis laevigata* Willd.

Ethyl acetate and water extracts of air-dried leaf material of *P. tomentosum* resulted in the isolation of the previously known flavonoids vitexin [3, 4], schaftoside (6-C-glucosyl-8-C-arabinosylapigenin) and isoschaftoside (6-C-arabinosyl-8-C-glucosylapigenin) [5] together with lesser amounts of apigenin 4'-O-glucoside and apigenin. Colour reactions in UV light before and after fuming with ammonia (olive) and spraying with Naturstoff reagent

(NA) (green) [6] are in accordance with those of apigenin derivatives. Isomerization with 0.1 N trifluoroacetic acid indicated the presence of C-glycosides. Cochromatography with authentic samples and UV,  $^1\text{H}$ - and  $^{13}\text{C}$  NMR data were in agreement with reported values [3, 7–9].

The present investigation has revealed that apigenin mono- and di-C-glycosides are the predominant compounds in *P. tomentosum*, the distributional pattern being uniform irrespective of the host plant. It is interesting to note that the methylated quercetin derivatives of *Viscum album* [2] also show a quite uniform pattern. However, further investigations on the flavonoids of other mistletoe species are needed before any conclusions on their value in systematic differentiation can be drawn.

#### EXPERIMENTAL

*P. tomentosum* leaves were collected by J. Exner near Austin, Texas, U.S.A. Voucher specimens are deposited in the Herbarium of the Botany Dept., University of Texas at Austin.

**Extraction and isolation** Air-dried leaves (200 g) were ground and extracted with 80%  $\text{MeOH}$  (1:1 × 3), filtered and evaporated

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to a syrupy residue under red pres at 30°. The syrup was partitioned between petrol (50–70°, 11), CHCl<sub>3</sub> (11), EtOAc (11) and H<sub>2</sub>O (11). EtOAc and H<sub>2</sub>O fractions containing flavonoids were further subjected to a Craig distribution with 70 steps in a two phase system as follows (1) EtOAc fraction, CHCl<sub>3</sub>–MeOH–PrOH–H<sub>2</sub>O (4.6:1.4, upper phase as mobile phase), and (2) H<sub>2</sub>O fraction, EtOAc–PrOH–H<sub>2</sub>O (4.3:5, upper phase as mobile phase). Individual fractions were monitored using Si gel rapid plates, Woelm F 254, using mobile phase (upper layer) as solvent system from the Craig two phase system. Flavonoid containing fractions were pooled and further chromatographed.

**Chromatography** CC cellulose microcrystalline, ashless quality, acid washed (solvents 0.05–0.1% HOAc, 10% MeOH), Sephadex LH 20 (solvent MeOH) TLC cellulose plastic sheets without fluorescent indicator (solvents 15% HOAc, 25% HOAc, BAW, upper phase), spray reagent NA.

**Hydrolysis and acid isomerization** 5 ml 0.1 N TFA [10] and 10 mg flavonoid heated in a steam bath for 1 hr. PC of sugars, Whatman 3 MM (solvent pyridine–EtOAc–HOAc–H<sub>2</sub>O, 36:36:7:1). Spray reagent Aniline hydrogen phthalate.

**UV spectroscopy** As described in ref [11].

**<sup>1</sup>H- and <sup>13</sup>C NMR spectroscopy** The spectra were recorded in DMSO-d<sub>6</sub> at 30°.

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running the <sup>1</sup>H- and <sup>13</sup>C NMR spectra. We also express our appreciation to Professor Zinsmeister for providing authentic samples for co-chromatography.

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## A FLAVANONE GLYCOSIDE FROM THE FRONDS OF *CETERACH OFFICINARUM*

FILIPPO IMPERATO

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(Received 12 May 1982)

**Key Word Index**—*Ceterach officinarum*, Polypodiaceae, naringenin 7-[O-L-arabinopyranosyl-(1 → 6) glucoside]

**Abstract**—From the fronds of the fern *Ceterach officinarum* naringenin and a new flavanone, naringenin 7-[O-L-arabinopyranosyl-(1 → 6) glucoside], have been characterized.

Early investigations of the chemical constituents of *Ceterach officinarum* Lam. et DC. have led to the identification of lignin [1], higher alkanes (the entire series from C<sub>19</sub> to C<sub>31</sub>) [2], triterpenoids [22 (29)-hopene and cyclolaudenol] [3] and by using neutron activation analysis, the sodium, potassium, chlorine and manganese levels have been determined in a pharmacological study [4] on diuretic drugs. Recently an examination of its polyphenolic constituents in this laboratory has led to the

identification of three hydroxycinnamic acid-sugar derivatives (1-caFFEYL glucose 6-sulphate, 1-caFFEYL glucose 3-sulphate and 1-caFFEYL glucose 2-sulphate) [5], and four flavonol glycosides quercetin 3-glucoside, quercetin 3-gentiobioside, kaempferol 3-(6"-malonyl) glucoside and kaempferol 3-(6"-malonyl) galactoside [6].

In the present study another flavonoid band was isolated from an ethanolic extract of fresh fronds of *C. officinarum*. The UV spectral data  $\lambda_{\text{max}}^{\text{MeOH}}$  nm 267 (sh), 282,